THE PREPARATION AND CHARACTERIZATION

OF [3H] HISTAMINE.2HC1 AT HIGH SPECIFIC ACTIVITY

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SUMMARY

[3 H] Histamine.2HCl will continue to play a key role in establishing the neurotransmitter status of histamine.2HCl. To more consistently achieve high specific activity in the preparation of the radioligand and incorporate as much tritium into the stable side chain positions as possible, we explored the use of alternative tritiation precursors. Imidazole 4-acetonitrile (5) and 2,5-diiodoimidazole-4-acetonitrile (6) were catalytically reduced with tritium gas and the radiolabeling specificity of the resulting [3 H] histamine.2HCl produced was confirmed by 3 H NMR.

Key Words: [³H] Histamine, Tritium Labeling, ³H NMR, Catalytic Reduction

INTRODUCTION

The physiological actions of histamine.2HCl (1) were described as early as 1910,² and histaminergic is the term applied to a certain population of receptors (H₁ and H₂) that regulate body temperature, gastric secretion and certain smooth muscle contraction. Although mounting neurochemical evidence supports the role of <u>1</u> as a neurotransmitter in mammalian brain,³, its status as such is still putative. To assist in firmly establishing the neurotransmitter role of <u>1</u>, [³H] histamine.2HCl will continue to be utilized in autoradiography, receptor binding assay⁴,⁵ and electrophysiological studies.⁶ [2-³H] Histamine.2HCl (2) at low specific activity has been prepared by general exchange tritiation of <u>1</u>,⁷ and [2,5-³H] histamine.2HCl (3) at high (25-50 Ci/mmol) specific activity has been prepared by the catalytic tritiation of 2,5-diiodohistamine (4).⁸ To more consistently achieve high specific activity

0362-4803/87/060615-08\$05.00 © 1987 by John Wiley & Sons, Ltd. Received July 29, 1986 Revised October 7, 1986 in the preparation of $[{}^{3}H]$ histamine.2HCl and incorporate as much tritium into the stable side chain positions as possible, we explored the use of nitriles <u>5</u> and 6 as alternative precursors.

DISCUSSION

Nitrile <u>5</u> is commercially available. Although <u>1</u> can easily be diiodinated to <u>4</u>,⁹ and histidine-HCl (<u>7</u>) can be diiodinated to 2,5-diiodohistidine-HCl (<u>8</u>)¹⁰, nitrile <u>5</u> could not be directly diiodinated to 2,5-diiodoimidazole-4-acetonitrile (<u>6</u>). A recent report¹¹ illustrated the difficulty and confusion that can attend the iodination of imidazoles. Exploiting the established procedure to prepare <u>5</u> from <u>7</u>,¹² we found that novel compound <u>6</u> could be prepared in 31% yield by the treatment of <u>8</u> with sodium hypochlorite.

Nitrile <u>5</u> was catalytically reduced with tritium and PdO in acetic anhydride followed by hydrolysis to yield [ring-2, ethanamine-³H] histamine.2HCl (9) at 44 Ci/mmol in 3.3% radiochemical yield. A ³H NMR (CD₃OD, DCl - Figure 1) of <u>9</u> revealed that 23% of the radiolabel was incorporated in and retained by the 2-position with the remainder residing in the side chain. Monitoring the ³H NMR of <u>9</u> in acidic solution at 4°C for several weeks showed that essentially no exchange of the 2-³H occurred. This general exchange of tritium into the 2-position of <u>9</u> is reminiscent of the result observed when <u>1</u> is exposed to tritium gas in the presence of 5% Rh/Al₂O₃ affording <u>2</u> as shown by ³H NMR.⁷

Hydrogenation of <u>6</u> with PdO in acetic anhydride did not yield any <u>1</u>. To facilitate both the dehalogenation and clean nitrile reduction of <u>6</u>, more basic conditions proved necessary. Exposure of <u>6</u> to tritium gas in EtOH:NH₄OH using both 10% Pd/C and 5% Rh/Al₂O₃ afforded [ring-5, ethanamine-³H] histamine.2HC1 (<u>10</u>) at 35 Ci/mmol in 17.3% radiochemical yield. A ³H NMR (CD₃OD, DCl - Figure 2) of <u>10</u> revealed that 42% of the radiolabel was incorporated in and retained by the 5-position with the remainder in the side chain. When compound <u>6</u> was stirred with 10% Pd/C and 5% Rh/Al₂O₃ in EtOH:NH₄OH for 24 h it was recovered unchanged. Therefore, the absence of tritium in the 2-position of compound <u>10</u> occurred as a result of exchange with the solvent after tritiation. To better understand the factors governing the rate of exchange at the 2-position of <u>10</u>, ²H NMR studies were conducted. [2,5-²H] Histamine-2HCl was prepared by the catalytic deuteration of <u>4</u> and treated with EtOH:NH₄OH with and without 10% Pd/C and 5% Rh/Al₂O₃. Also, <u>1</u> was treated with EtOD:ND₄OD with and without 10% Pd/C and 5% Rh/Al₂O₃. In these parallel experiments it was observed by ²H NMR that both catalyst and base were necessary for a facile exchange of the 2-position.

Both compounds <u>9</u> and <u>10</u> were purified by ascending paper chromatography and were found to be 98% radiochemically pure by TLC and HPLC, and also cochromatographed with authentic <u>1</u>. The specific activity of <u>9</u> and <u>10</u> was determined by fluorescamine assay. Nitriles <u>5</u> and <u>6</u> have been shown to serve as useful precursors to $[^{3}H]$ histamine-2HCl at high specific activity, incorporating a majority of the radiolabel in the stable side chain position.

EXPERIMENTAL PROCEDURES

General Methods: Evaporations were carried out on a Buchi rotary evaporator. Analytical TLC was performed either on Analtech 5 x 15 cm silica gel or avicel GF coated glass plates. Common solvent combinations were S₁ (CHCl₃:CH₃OH (9:1)); S_2 (PhH:acetone (9:1)); S_3 (PhH:CHCl₃:EtOH (2:4:1)); S_4 (EtOH:NH_OH (4:1)); S₅ (n-BuOH:HOAc:H₂O (25:4:10)). Autoradiography was performed at O°C after spraying TLC plates with PPO (DuPont, NEN Research Products) and exposure to Eastman Kodak SB-5 film. The plates were also scanned tor radioactivity by using a Packard 7201 scanner. Analytical HPLC determinations were run on a Waters instrument using a μ C₁₈ column. Common solvent combinations were S_6 (0.01N KH_2PO_4 (pH = 2.5):CH₃OH (70:30)); S_7 (Pic B7:CH₂OH (70:30)). Peak detection was performed simultaneously by a liquid scintillation flow monitor and UV (254 nm) detection. The IR spectrum was measured on a Perkin-Elmer Model 700 spectrophotometer. The proton and triton magnetic resonance spectra were obtained on a Bruker WP 200 mHz NMR spectrometer. Chemical shift values are expressed in parts per million downfield form internal TMS or TSP. Fluorescence measurements were determined using a Turner Model 430 fluorimeter. The UV spectrum was measured on a Beckman model 25 spectrophotometer and the high resolution mass spectrum was performed by Shrader Analytical Laboratories (Detroit, MI).

2,5-Diiodoimidazole-4-acetonitrile (6). To a suspension of

L-2,5-diiodohistidine. HCl¹⁰ (<u>8</u>, 900 mg, 2.03 mmol) in 40 mL of H₂O at 0°C was added with vigorous stirring 6.30 mL of a 5.25% sodium hypochlorite solution (Chlorox) dropwise over the course of several min. Gas (CO₂) evolution was noted immediately. The reaction was allowed to warm to room temperature and stir overnight. After this time, the dark acidic solution was basified to pH 9 with excess Na₂CO₃ and cautiously evaporated to dryness (foaming). The residue was slurried with twenty 20 mL portions of EtOAc. The combined EtOAc washings were filtered and evaporated to yield 550 mg of a residue. The residue was preparatively chromatographed on six 1000 µm silica gel GF plates developed with S₁. The main band (R_f 0.7) was visualized by UV, scraped and eluted with EtOAc. Solvent evaporation yielded 225 mg (31%) of <u>6</u> as a white solid, m.p. 173-175°C, that was found to be homogeneous on TLC (Silica Gel - S₁, S₂, S₃) and HPLC (µ C₁₈ - S₆) ¹H NMR (CD₃OD) δ 3.77 ppm (s); ¹³C NMR (CD₃OD) 117.46, 87.95, 79.49, 16.75 ppm; IR (KBr) 3080, 3000, 2900, 2820, 2650, 2270, 1555, 1475, 1400, 1050, 980 cm⁻¹; UV (EtOH) ϵ 230 (10,840).

High resolution mass spectrum calcd for $C_5H_3N_3I_2$ (M⁺):358.8421. Found 358.8416.

[Ring-2, Ethanamine-³H] Histamine·2HCl (9). A solution of

imidazole-4-acetonitrile ($\underline{5}$, 11 mg, 0.1 mmol) in 3 mL of acetic anhydride with 25 mg of PdO was reduced with 70 Ci of tritium gas for 16 h at 24°C. Following catalyst filtration and removal of labile tritium, the residue was dissolved in 5 mL of 2N HCl and heated at 100°C for 16 h. Following solvent evaporation and removal of labile tritium with CH₃OH, the residue was dissolved in 10 mL CH₃OH:H₂O (1:1). The crude product (4015 mCi) was purified twice by ascending paper chromatography; first on three Whatman 3 mm sheets followed by purification on two Whatman 1 mm sheets developed each time overnight with i-PrOH:Conc·HCl:H₂O (26:8:6). To facilitate product location, cold histamine·2 HCl (1) was allowed to migrate at the side of each paper. After each overnight development, the papers were dried, autoradiogrammed, and the main radioactive band lining up with visualized (ninhydrin) cold standard histamine ·2HCl (R_f 0.5) was cut out and eluted with EtOH:O.01 N HCl (1:1) affording 146 mCi(3.3%



Scheme 1



Figure 1. 3 H NMR (CD₃OD, DCl) of [ring-2,ethanamine- 3 H] histamine· 2 HCl (2). Chemical shift values are in parts per million from internal TSP.



Figure 2. 3 H NMR (CD₃OD, DCl) of [ring-5,ethanamine- 3 H] histamine-2 HCl (<u>10</u>). Chemical shift values are in parts per million from internal TSP.

radiochemical yield based on <u>5</u>) of <u>9</u>. Ligand <u>9</u> was found to be 98% radiochemically pure by TLC (silica gel - S_4 , avicel - S_5) and HPLC (μ C₁₈ - S_7) and in these systems <u>9</u> cochromatographed with authentic <u>1</u>. A specific activity (fluorescamine) of 44 Ci/mmol was determined for <u>9</u>. A ³H NMR (CD₃OD,DCl) of <u>9</u> (Figure 1) afforded a singlet at δ 8.72 ppm and a multiplet at δ 3.40 -3.00 ppm.

[Ring-5, Ethanamine-³H] Histamine-2HCl (10). A solution of 2,5-diiodoimidazole-4-acetonitrile (6, 32 mg, 0.09 mmol) in 6 mL of EtOH:NH₄OH (2:1) with 20 mg each of 10% Pd/C and 5% $\rm Rh/Al_2O_3$ catalyst was reduced with 80 Ci of tritium gas for 16 h at 24°C. Following catalyst filtration and removal of labile tritium, the residue was dissolved in 10 mL $CH_2OH:H_2O$ (1:1). The crude product (1548 mCi) was purified by ascending paper chromatography on two Whatman 3 mm sheets developed overnight with i-PrOH:conc. HC1:H₂O (26:8:6). To facilitate product location, cold histamine \cdot 2HCl (1) was allowed to migrate at the side of each paper. After overnight development the papers were dried autoradiogrammed and the main radioactive band lining up with visualized (ninhydrin) cold standard histamine-2HCl (R_f 0.5) was cut out and eluted with EtOH:0.01 N HCl (1:1) affording 540 mCi (17.3% radiochemical yield based on 6) of 10. Ligand 10 was found to be 98% radiochemically pure by TLC (silica gel - S_4 , avice1 - S_5) and HPLC (μ C_{18} - S_7) and in these systems <u>10</u> cochromatographed with authentic <u>1</u>. A specific activity (fluorescamine) of 35 Ci/mmol was determined for <u>10</u>. A 3 H NMR (CD₂OD, DC1) of <u>10</u> (Figure 2) afforded a singlet at δ 7.47 ppm and a multiplet at δ 3.35 - 3.10 ppm.

ACKNOWLEDGEMENT

We gratefully acknowledge the technical assistance of K. Bradley in the aforementioned tritiations and the help of Dr. P. Srinivasan and L. Thomas in obtaining the 1 H, 2 H, 3 H and 13 C NMR spectra. We also thank M. Tutunjian and R. Wellman for analytical HPLC work.

REFERENCES

 Presented in part at the 12th Northeast Regional Meeting of the American Chemical Society, Burlington, Vermont, June, 1982, Abstract #253.

- Dale H.H. and Laidlaw P.P. J. Physiol. (London). <u>41</u>: 318 (1910).
- 3. Schwartz J.C., Pollard H. and Quach T.T. J. Neurochem. 35: 26 (1980).
- Batzri S., Harmon J.W. and Walker M.D. Biochem. Biophys. Res. Commun. 108, 965 (1982).
- 5. Batzri S. Biochem. Pharmacol. <u>30</u>: 3013 (1981).
- 6. Subramanian N. Life Sciences <u>31</u>: 557 (1982).
- 7. Filer C.N. and Ahern D.G. unpublished results.
- Bloxsidge J.P., Elvidge J.A., Gower M., Jones J.R., Evans E.A., Kitcher J.P. and Warrell D.C. - J. Labelled Compd. Radiopharm. <u>18</u>: 1141 (1981).
- 9. Tominaga M. and Paiva A.C.M. J. Med. Chem. <u>12</u>: 693 (1969).
- 10. Brunings K.J. J. Amer. Chem. Soc. 69: 205 (1947).
- Dickens J.P., Dyer R.L., Hamill B.J., Harrow T.A., Bible R.H., Finnegan P.M., Henrick K. and Owston P.G. - J. Org. Chem. <u>46</u>: 1781 (1981).
- 12. Bauer H. and Tabor H. Biochem. Prep. 5: 97 (1957)